STEREOSPECIFIC TRANSFORMATION OF D-SUGAR TO L-SUGAR IN COMPLEX AMINOGLYCOSIDE. SYNTHESIS OF A KANAMYCIN B ANALOG HAVING 2,6-DIAMINO-2,4,6-TRIDEOXY-L-ARABINO-HEXOPYRANOSE

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Abstract: A 4'-ene derivative of kanamycin B (<u>4</u>) was derived from the epoxide (<u>1</u>) by oxidative elimination of the 4'-phenylseleno group into the allylic alcohol (<u>3</u>). The title compound, $O-(2,6-\text{diamino}-2,4,6-\text{trideoxy}-\beta-\text{L-arabino}-\text{hexopyranosyl})-(1-+4)-O-[3-\text{amino}-3-\text{deoxy}-\alpha-D-gluco-pyranosyl-(1-+6)]-2-deoxystreptamine (<u>6</u>) was obtained from <u>4</u> by stereospecific hydrogenation followed by removal of the masking groups, changing the D-sugar molety of the 4-O-glycoside portion into an L-sugar.$

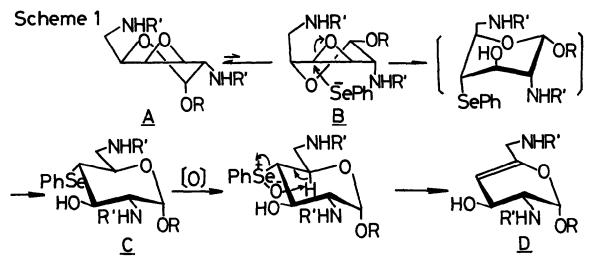
In order to develop compounds useful in the treatment of resistant infections, extensive studies have been continued in chemical derivation of kanamycin and structure-activity relation-ships¹⁻³⁾. However, little is known concerning the conformational change of the ring A except for the altered conformations of the ring A due to the 3',4'- or 4',5'-double bond formation as exemplified by 3',4'-unsaturated kanamycin B¹⁾ and sisomicin⁴⁾. The present paper describes a new modification of kanamycin B by the transformation of the ring A (D-sugar) into the L-form. The synthesis involves a novel formation⁵⁾ of 4',5'-double bond in the ring A and its dramatic transformation into L-hexopyranoside (ring A') by stereospecific hydrogenation.

For the formation of 4,5-double bond in the ring A it was designed to convert the epoxide (<u>A</u>) into the allylic alcohol (<u>D</u>) by oxidative elimination of 4'-phenylseleno group⁶) as shown in Scheme 1. The trans-diequatorial opening of the epoxide was successful by the use of a bulky nucleophile (PhSe⁻) to give <u>C</u> as expected from literatures^{6,7)}. The attack of the bulky phenyl-seleno anion at C-3 is hindered by the anomeric axial group in <u>A</u>, and the less stable conformer <u>B</u> is attacked at C-4 position. An epoxide derivative⁸ (<u>1</u>) of kanamycin B was chosen as the starting material.

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Treatment of 1 with sodium phenylselenide generated in situ (PhSeSePh, NaBH₄, dimethoxyethane, 60°C) gave 4'-phenylseleno derivative (2) in 68% yield, $[\alpha]_D^{20}$ +58.5° (c 0.8, CHCl₃). The regio- and stereochemistry of C-3' and C-4' of 2 were best assigned in the deblocked derivative (7)^{9,10} and in the reductive conversion to known 4'-deoxykanamycin B¹¹). The derivative 7 was derived from 2 by treatment with sodium methoxide in methanol and then with 50% aqueous trifluoroacetic acid. The deoxy derivative 8 was obtained from 7 by catalytic hydrogenation with Raney Ni. 7: $[\alpha]_D^{20}$ +81.8° (c 0.92, H₂0); PMR (D₂0) δ 5.78 (1H d, J=4.0 Hz, H-1'), 5.33 (1H d, J=4.0 Hz, H-1"); ¹³CNMR (D₂0) δ 100.8 (C-1'), 99.7 (C-1"), 72.9 (C-3'), 57.6 (C-4'). 8: $[\alpha]_D^{20}$ +125° (c 1.0, H₂0) [Lit.¹¹ +122° (c 0.25, H₂0)]; ¹³CNMR (D₂0) δ 101.9 (C-1'), 100.7 (C-1"), 36.81 (C-4'). ¹³CMR spectrum of 7 shows 13.6 ppm up-field shift for C-4' at δ 57.6 compared with that of kanamycin B at δ 71.2¹², clearly indicating the presence of phenylseleno group at C-4'. All spectral means (PMR, ¹³CNMR, IR) and specific rotation of 8 were superimposable to those of authentic sample of 4'-deoxykanamycin B¹¹, indicating the equatorial hydroxy group at C-3'.

Oxidation of <u>2</u> with m-chloroperbenzoic acid (mCPBA) followed by thermal elimination⁶⁾ (mCPBA, 4-dimethylaminopyridine, CH₂Cl₂, room temperature, and then reflux in toluene) produced the allylic alcohol (<u>3</u>)^{9,10)} in 60.6%: $[\alpha]_D^{20}$ +84.5° (c 1.4, CHCl₃); ¹³CNMR (CDCl₃) δ 147.7 (C-5'), 100 (C-1', C-1"), 101.4 (C-4'); PMR (CDCl₃) δ 5.6 (1H broad s, H-4').

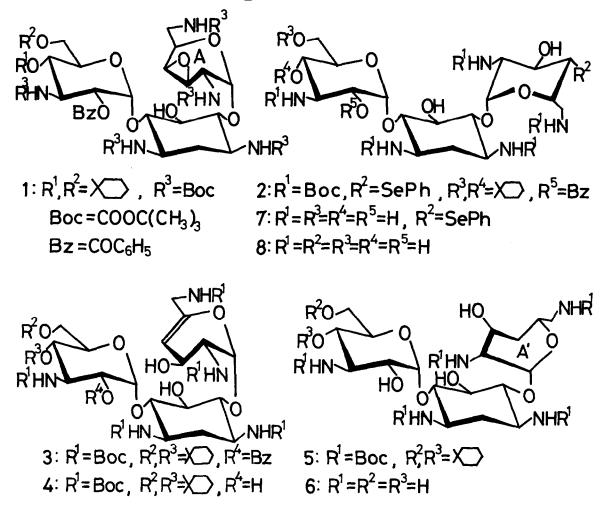
Debenzoylation of <u>3</u> with sodium methoxide in methanol gave the triol $(\underline{4})^{9,10}$ in 99%: $[\alpha]_{D}^{20}$ +50° (c 1.0, CHCl₃); ¹³CNMR (CDCl₃) &147.5 (C-5'), 101.4 (C-4'), 100 (C-1', C-1''). Stereospecific hydrogenation¹³⁾ of <u>4</u> was best achieved by catalytic hydrogenation with

Stereospecific hydrogenation¹³⁾ of <u>4</u> was best achieved by catalytic hydrogenation with Adams' catalyst in ethanol at room temperature under atmospheric pressure to give $5^{9,10}$ in 89%: $\left[\alpha\right]_{D}^{20}$ +35.7° (c 0.84, CHCl₃). ¹³CNMR spectrum of <u>5</u> showed no peak around δ 147.5 and 101.4, indicating that <u>5</u> has no corresponding double bond.

Finally, removal of the t-butoxycarbonyl and cyclohexylidene groups by treatment with 50% aqueous trifluoroacetic acid at room temperature afforded 0-(2,6-diamino-2,4,6-trideoxy- β -L-arabino-hexopyranosyl)-(1- 4)-0-[3-amino-3-deoxy- α -D-glucopyranosyl-(1- 6)]-2-deoxystreptamine (<u>6</u>) in 77%: $[\alpha]_D^{20}$ +94.5° (c 0.75, H₂0); ¹³CNMR (D₂0) &101.0 (C-1', C-1''), 72.7 (C-5''), 30.01

(C-4'); PMR (D₂0) $\delta 5.40$ (1H d, J_{1',2'}=1.5 Hz, H-1'), 5.36 (1H d, J_{1'',2''}=4.0 Hz, H-1''), 3.29 (1H dd, J_{1',2'}=1.5, J_{2',3'}=3.5 Hz, H-2'). ¹³CNMR spectrum of <u>6</u> showed ~ 41 ppm up-field shift for C-4' at $\delta 30.06$ compared with that of kanamycin B at $\delta 71.2^{12}$, clearly indicating the methylene carbon at C-4'. The small coupling constants of H-1' (d, J_{1',2'}=1.5 Hz) and H-2' (dd, J_{1',2'}=1.5 and J_{2',3'}=3.5 Hz) suggested that the conformation of 4-0-glycoside portion (ring A') was a skew boat (B_{4,1}).

The semisynthetic aminogly coside $\underline{6}$ was found to have a weak antibacterial activity.



Acknowledgements: We wish to thank Dr. H. Naganawa, Institute of Microbial Chemistry, for PMR spectra and Dr. E. Akita, Central Research Laboratories, Meiji Seika Kaisha Ltd., for his help-ful advices and encouragement.

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- 9) This compound has been fully characterized by spectral means and elemental composition determined by mass spectroscopy and elemental analysis.
- 10) <u>3</u>: ¹³CNMR δ166.21, 157.37, 155.41, 155.23, 147.66, 133.51, 130.10, 129.20, 128.58, 101.35, 100.01, 99.08, 97.86. <u>4</u>: ¹³CNMR δ157.61, 156.15, 155.39, 147.50, 101.41, 100.03, 99.78, 99.29; mass spectrum (FD) m/e 1045. <u>5</u>: ¹³CNMR δ154.62, 154.33, 153.96, 153.64, 97.44, 96.92, 96.67. <u>6</u>: 100 MHz-NMR δ5.40 (1H d, J=1.5 Hz), 5.36 (1H d, J=4.0 Hz), 3.83 (1H dd, J=4.0, 10 Hz), 3.29 (1H dd, J=1.5, 3.5 Hz), 2.31 (1H dt, J=4.0, 13 Hz), 1.4-2.1 (3H m); ¹³CNMR δ100.96, 88.47, 87.18, 74.96, 73.44, 72.69, 70.03, 61.01, 55.03, 53.26, 51.52, 49.97, 45.71, 36.28, 30.06. <u>7</u>: 100 MHz-NMR δ8.05 (2H m), 7.78 (3H m), 5.78 (1H d, J=4.0 Hz), 5.32 (1H d, J=4.0 Hz); ¹³CNMR δ138.08, 130.34, 130.26, 124.78, 100.76, 99.67, 88.97, 84.08, 75.66, 72.93, 72.58, 70.29, 70.08, 61.14, 57.56, 55.04, 51.14, 50.15, 48.10, 43.82, 35.92. <u>8</u>: 80 MHz-NMR δ5.65 (1H d, J=4.0 Hz), 5.35 (1H d, J=4.0 Hz); ¹³CNMR δ101.89, 100.70, 88.75, 87.27, 75.35, 72.92, 72.64, 70.70, 70.14, 68.89, 61.18, 57.64, 55.05, 51.18, 50.16, 45.42, 36.81, 36.31.
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(Received in Japan 15 September 1980)